

Advances in Sciences and Arts Reviews

Journal homepage:<https://asa.must.ac.mw/>

REVIEW ARTICLE

Category Science

***Correspondence to:**

cmalambabanda@must.ac.mw

Citations

Malamba-Banda C., et al (2023). T Cell Subsets and COVID-19: Function and Dysregulation Effect. *Advances in Sciences and Arts*. 1(1), T011-0001, https://doi.org/10.37872/ T011- 0001

Supporting info

These can be found on the journal's official website on <https://asa.must.ac.mw/>

Received 12 th Dec 2022

Accepted 18 th Feb 2023

Published 13 th Nov 2023

DOI

doi.org/10.37872/ T011-0001

T Cell Subsets and COVID-19: Function and Dysregulation Effect

C. Malamba-Banda^{1,2,3,4}, C. Kafamau¹, T. Nthandira¹, A. Kumwenda¹, D. Mzinza^{1,2}, G. Bandawe¹, W. Mandala^{1,3}

¹Malawi University of Science and Technology, Department of Biological Sciences ²Malawi Liverpool Welcome Trust Research Clinical Program, Virology research group

³Kamuzu University of Health Science, Department of Medical Laboratory Science ⁴University of Liverpool, Institute of Veterinary and Ecological Sciences

Abstract T-cell responses are an important component of the human immune response to viral infections and specifically for viral clearance, establishing memory recall and inducing antibody responses. In SARS-CoV-2 infected individuals, lymphopenia and/or T-cell exhaustion somehow contributes towards the development of severe diseases. In contrast, long-lived memory T cells play a key role in preventing severe cases of COVID-19 disease. Exposure to SARS-CoV-2 either through infection or vaccination generates immune cells that provide long-term immunity to SARS-CoV-2 which is crucial for the development of herd immunity. In this review, we summarize what is currently known about various T cell subsets such as CD4+ and CD8+ T cells in response to SARS-CoV-2 and the specific T cell responses linked to natural, vaccine-induced and hybrid immunity highlighting the need to look beyond blood to fully understand how T cells function in the tissue. Understanding the T cell-specific immune responses in various body tissues could facilitate the formulation of better therapeutic strategies for COVID-19 patients.

Keywords: *Cell-mediated immunity; SARS-CoV-2; Immune responses, T helper cells; Cytotoxic T cells; Adaptive immunity*

Introduction

Coronavirus disease 2019 (COVID-19) is caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) strain (Wu et al., 2020; P. Zhou et al., 2020). This pandemic first surfaced in December 2019 in Wuhan, China and spread worldwide (Wu et al., 2020; P. Zhou et al., 2020). SARS-CoV-2 contains four structural proteins:

spike (S) protein, envelope (E) protein, membrane (M) protein, and Nucleocapsid (N) protein and sixteen non-structural proteins (NSP1 to 16) (Naqvi et al., 2020; M. Y. Wang et al., 2020). Coronaviruses' entry into host cells is mediated by the S protein (M. Y. Wang et al., 2020). SARS-CoV-2 enters human cells via receptor angiotensinconverting-enzyme 2 (ACE2) and the co-receptor transmembrane serine protease 2 (TMPRSS2) (Scudellari, Megan, 2021; Walls et al., 2020; Wrapp et al., 2020). SARS-CoV-2 initiates both innate and adaptive immune responses (García, 2020). Early immune responses primarily have a protective role, however, dysregulated and exacerbated inflammatory responses can fail in viral clearance and lead to worse disease outcomes (García, 2020; Toor et al., 2021). T cells form a key component of the adaptive immune response to viral infection (Toor et al., 2021). SARS-CoV-2 elicits antiviral immune responses by helper T cells (Th) and cytotoxic T cells (CTLs) (Toor et al., 2021). Antiviral immunity is mediated by the generation of neutralizing antibodies generated by B cells and CTL-mediated immunity (García, 2020; Toor et al., 2021). Cytokines produced by Th cells help immune cell recruitment and activate both CTLs and B cells. CTLs are important because of their specific cytotoxicity for infected cells.

SARS-CoV-2 can infect T cells via S proteinmediated endocytosis and internalization although the virus is not able to replicate in these cells (W. Wang et al., 2020). A decline in peripheral T cell subsets is a unique characteristic in patients presenting with acute SARS-CoV-2 infection and this is followed by a rapid normalization of the cell counts during the convalescence (T. Li et al., 2004). As demonstrated for the Middle East Respiratory Syndrome (MERS) coronavirus, the translation of viral RNA proteins induces apoptosis in T cells, leading to the lymphocytopenia observed in COVID-19 patients (Chu et al., 2016). Lymphopenia and/or T-cell exhaustion can worsen clinical outcomes in COVID-19 patients, whereas T-cell-mediated inflammation can contribute towards lung pathology and secondary complications seen in severe disease cases (García, 2020).

2. Materials and Methods

2.1 Literature Search Strategy

This study is a scoping review. We searched on Pubmed for literature on:

- 1. What are the types of T cells that are associated with COVID-19 disease caused by SARS-CoV-2?
- 2. What are the phenotypes of the SARS-CoV-2 specific T cells during and following COVID-19 disease?
- 3. What are the dysregulation effects of the T cells induced following COVID-19 disease caused by SARS-CoV-2?

We used a search strategy composed of a combination of the following search terms: "T cells and COVID-19", "T cell immune response and COVID-19" "T cell function in COVID-19 disease", "CD4+/CD8+ in SARS-CoV-2 infection", "SARS-CoV-2 specific T cells", "SARS-CoV-2 specific T cells and diabetes", "SARS-CoV-2 specific T cells and Tuberculosis".

2.2 Inclusion and Exclusion

The study included articles that targeted both animal and human participants. These would be articles which contained information answering our research questions. The most important is that it should be clear and sufficient information, including positive or negative, to answer the questions. We excluded mostly the unrelated, duplicated, unavailable full texts, or abstract-only papers. These exclusions were stated in advance to refrain the researchers from bias.

3. Results and Discussion

3.1 T helper (Th) cells

T helper (CD4+) cells have been associated with the control of primary SARS-CoV-2 with the SARS-CoV-2-specific CD4+ T cells thought to differentiate into T follicular helper (Tfh) and Th1 cells (Or Caspi, Michael J. Smart, 2020). SARS-CoV-2-specific CD4+ T cells predominantly have a central memory (CD45RA-, CCR7+) phenotype (Spoerl et al., 2021) and central memory cells have the potential to home secondary lymphoid tissue (Martin et al., 2015). Tfh cells provide help to B cells for affinity maturation, antibody production and long-term humoral immunity and or memory B cells.

SARS-CoV-2-specific circulating Tfh (cTfh) frequencies have been associated with disease severity and a substantial fraction of SARS-CoV-2 cTfh are CCR6+ (Juno et al., 2020). CCR6 is a chemokine receptor associated with migration to mucosal tissues (Weiskopf et al., 2020a). Potentially cTfh that are CCR6+ would be indicative of mucosal airway homing as observed in common cold coronavirus HKU1 (Weiskopf et al., 2020a). Expression of CCR6 may also indicate the existence of Th17 cells however the expression of interleukin-17α (IL-17α) protein, the marker for Th17 cells on these cells has been reported to be low or undetectable (Or Caspi, Michael J. Smart, 2020; Rydyznski Moderbacher et al., 2020). SARS-CoV-2-specific CD4+T cells produce IL-22 robustly which is associated with tissue repair in the lung (Dudakov et al., 2015; Rydyznski Moderbacher et al., 2020) and gut suggesting that SARS-CoV-2 specific CD4+ T cells may actively participate in lung tissue repair during COVID-19 (Dudakov et al., 2015; Rydyznski Moderbacher et al., 2020).

The cells have antiviral activities which are elicited through the production of interferon-gamma (IFNγ) (Or Caspi, Michael J. Smart, 2020; Rydyznski Moderbacher et al., 2020; Weiskopf et al., 2020a) and other pro-inflammatory cytokines. The dominant cytokine produced by SARS-CoV-2 specific CD4+ T cells from COVID-19 patients is IFN-γ (Or Caspi, Michael J. Smart, 2020; Rydyznski Moderbacher et al., 2020; Weiskopf et al., 2020a). SARS-CoV-2-specific CD4+ T cells from patients with COVID-19 consistently do not have Th2 characteristics (Grifoni et al., 2020) but in other few COVID-19 patients, Th2 responses

P a g e | **3**

were prevalent (Grifoni et al., 2020). CD4+ T cells, assist cytotoxic (CD8+) T cells to proliferate and differentiate. CD4+ CTL can have direct cytotoxic activity against virally infected cells in a class II antigen presentation-restricted manner although the cytotoxicity degranulation marker CD107a has been minimally observed on SARS-CoV-2 specific CD4+ T cells (Or Caspi, Michael J. Smart, 2020; Weiskopf et al., 2020a). The degranulation process is when the granule membrane is fused with the plasma membrane of the activated CD8+ T-cell, and the content of the granule is released into the immunological synapse between the CD8+ T-cell and the target cell (Or Caspi, Michael J. Smart, 2020; Weiskopf et al., 2020a).

Some studies have reported that COVID-19 patients have cytokine profiles of IL-2, IFN-γ, and IL-4 (Akbari et al., 2020; Gadotti et al., 2020). T cell response revealed SARS-COV-2 specific CD4+ IL-2-producing CD4+ in COVID-19 patients, in up to 75% of the patients displaying bior trifunctional INF-γ/IL-2/TNF-α profile (Tincati et al., 2020). Similar SARS-COV-2 specific CD8+ functionality was determined despite lower frequency and magnitude. The polyfunctional Tcell responses have been associated with better immunity in several infections (Harari et al., 2006; Oh et al., 2012, 2019; Shin et al., 2019).

Regulatory T (Treg) cells are a subset of CD4+ T cells involved in maintaining immune homeostasis and self-tolerance by inhibiting the proinflammatory activities of CD4+ and CD8+ effector T cells, natural killer cells, and antigenpresenting cells (M. O. Li & Rudensky, 2016; Newton et al., 2016; van der Veeken et al., 2016). Naturally occurring Treg cells are CD4+, CD25+, and Forkhead box P3 (FoxP3)+ cells that develop in the thymus (Schmetterer et al., 2012). The role of Treg cells during viral infection is for the maintenance of immune homeostasis, specifically, in controlling the severity of viral immunoinflammatory lesions (Suvas et al., 2004). Pattern changes in the frequency of Treg cells in

COVID‐19 patients have been observed and this could be one of the reasons for the hyperactivated immune responses and damaged lungs in severe COVID‐19 patients. Tregs may play an important role in the regulation of immune responses to COVID-19 (H. Wang et al., 2021). Treg cells can prevent cytokine storms, accelerate the resolution of acute respiratory distress syndrome (ARDS), and suppress the development of lung inflammatory disorders during the COVID-19 disease (Lee et al., 2010; Loebbermann et al., 2012; Rahimzadeh & Naderi, 2021).

3.2 Cytolytic (CTL) cells

CTLs mostly CD8+ T cells are very important in clearing the virus-infected cells. The presence of virus-specific CD8+T cells has been associated with better COVID-19 infection outcomes (Jordan, 2021). SARS-CoV-2 CD8+ T cells are specific for a range of SARS-CoV-2 antigens, with S, N, M, and open reading frames (ORF3a) proteins well represented. (Jordan, 2021). In acute COVID-19, SARS-CoV-2-specific CD8+ T cells exhibit high levels of molecules associated with potent cytotoxic effector functions, such as IFN-γ, granzyme B, perforin, and CD107 (Rydyznski Moderbacher et al., 2020). Activated CD8+ T cells kill virus-infected cells using TNF-related apoptosis-inducing ligand, secretion of cytolytic molecules (perforin and granzymes) and the binding of CD8+ T FasL to Fas expressed on the surface of virus-infected cells thereby initiating death-receptor pathway (Jiang et al., 2020).

CD8+ T cells in COVID-19 patients highly express CD38, human leukocyte antigen-DR isotype (HLA-DR), CD69, CD25, CD44 and Ki67 confirming that CD8+ T cells during the SARS-CoV-2 infection are highly activated and are capable of proliferation (Jiang et al., 2020). SARS-CoV-2–specific CD8+ T cells include large proportions of both central memory (CD45RA-, CCR7+) and terminally differentiated cells (CD45RA+, CCR7-) (Spoerl et al., 2021). Central memory T cells have greater recall potential,

enhanced IL2 production, proliferation, and survival capacity (Martin et al., 2015). Central memory T cells can differentiate into either Th1 or Th2 in the absence of polarizing cytokines (Martin et al., 2015). Terminally differentiated T cells reexpress CD45RA and display a loss in multipotency. Multipotency is the ability of the cells to become another cell type and to proliferate (Weissman, 2000).

3.3 Evidence for a protective role of T-cell immunity in the control of COVID-19 infection

There is increasing evidence that cellular immunity supports have a potential role in both preventing initial SARS-CoV-2 infection and limiting the magnitude of disease following infection (Moss, 2022; Sette & Crotty, 2021). The adaptive immune response is required for viral clearance during COVID-19 infection. Antibodies play an important role in viral neutralisation, however, there is evidence that the virus can spread through cell-tocell contact, which is immune to antibody neutralisation (Zeng et al., 2022). Several other viruses have demonstrated this mechanism (Skelly et al., 2021) suggesting that T-cell-mediated immunity may be important for viral clearance. Some reports have suggested that SARS-CoV-2 infection may induce T cells without seroconversion (Sekine et al., 2020a). Virusspecific memory T cells have been shown to persist for many years after infection with SARS-CoV-1 (Skelly et al., 2021). Memory CD4+ T cells and CD8+ T cells were detected in 100% and 70% of patients who recovered, respectively. These memory T cell responses were detected for multiple SARS-CoV-2 proteins, including spike protein as well as nucleoprotein and membrane protein.

SARS-CoV-2-specific T cells in the convalescent COVID-19 infection phase were required to acquire an early differentiated memory phenotype (CCR7+ CD127+ CD45RA−/+ T cell factor 1 (TCF1+)). This phenotype has been associated with stem-like properties and was previously reported in the context of other viral infections and successful vaccines (Neidleman et al., 2020). In the convalescent phase, SARS-CoV-2-specific T cells showed anamnestic responses to cognate antigens, marked by substantial proliferation and polyfunctionality (Sekine et al., 2020b). These identified memory T cell responses were directed against both the internal (nucleocapsid) and surface proteins (membrane and/or spike) in some individuals lacking detectable circulating antibodies specific for SARS-CoV-2. Further, almost twice as many healthy individuals who donated blood during the pandemic had memory SARS-CoV-2 specific T cell responses versus antibody responses (Sekine et al., 2020b). In another study, they found that SARS-CoV-2 reactive T cells in convalescent patients acquired a multifunctional (triple positive for INF-γ, IL-2 and TNFα) phenotype, which may be considered a correlate of protective immunity (Seder et al., 2008; Steiner et al., 2020).

The current COVID-19 vaccines elicit robust Tcell responses that likely contribute to protection against hospitalization or severe disease leading to death. A robust, stable, and fully functional spike protein-specific CD8+ T cell responses were elicited after prime mRNA vaccination in individuals who had low levels of neutralizing antibodies. (Oberhardt et al., 2021). In the same study following vaccination, the proportion of naive spike protein-specific CD4+ T cells decreased, suggesting a vaccine-induced activation (Oberhardt et al., 2021). T-cell responses are also important for generating and maintaining highaffinity antibodies. Dual vaccination with BNT162b2 led to reliable induction of virusspecific CD4+^T cell responses (Papadatou et al., 2023). These CD4+ responses detected by day 8 after priming expressed high levels of IFN-γ and IL-2, and low levels of IL-4, indicating a Th1 cell profile (Skelly et al., 2021). T-cell responses after dual vaccination are of similar magnitude to those seen after natural infection, although they seem to be somewhat more differentiated (Sahin et al., 2021).

T cell cross-reactivity is a phenomenon of the immune system defined as the recognition of two or more peptide-MHC complexes (pMHCs) by the T cell receptor (TCR) (Petrova et al., 2012). Crossreactivities of T cells against SARS-CoV-2 have been proposed and identified in patients with COVID-19 infections (Weiskopf et al., 2020b) by the presence of SARS-CoV-2 cross-reactive CD4+ T cells specific to the spike protein in donors who were not exposed to SARS-CoV-2 (Braun et al., 2020; Petrova et al., 2012). However, the biological role of the pre-existing cross-reactive CD4+ T cell remains unclear. Nevertheless, cellular responses induced by vaccines have shown strong crossprotection against variant concerns and support the concept that cellular responses contribute substantially to disease control (Skelly et al., 2021).

Hybrid immunity applies to individuals with a prior SARS-CoV-2 infection who were then vaccinated against SARS-CoV-2 or vice versa (Abu-Raddad et al., 2021; Bates et al., 2022; Goldberg et al., 2021; Kojima et al., 2021). It has been demonstrated that there is relatively higher protection for previously infected individuals with or without an additional vaccination dose compared to previously uninfected vaccinated individuals (Abu-Raddad et al., 2021). After several months persons with hybrid immunity are better protected against further infection compared to uninfected persons who have received two vaccine doses (Abu-Raddad et al., 2021). Data on the efficacy of hybrid immunity are inconsistent but point in the direction of hybrid immunity being superior as compared to either vaccine-induced (without a booster) or natural immunity alone (Abu-Raddad et al., 2021; Y. Wang et al., 2021). The protective importance of hybrid immunity is the combination of higher numbers of SARS-CoV-2-specific memory B cells, higher neutralizing antibody titers and the infection imprinted IFN- γ and IL-10 cytokine profile in CD4+ T cells (Y. Wang et al., 2021).

3.4 Dysregulation of T cells in COVID-19 patients

Although T cells help to coordinate antiviral immune responses, support humoral immune responses, limit viral replication, and remove infected cells, some studies have reported an exhausted or senescent T cell phenotype with decreased polyfunctionality and cytotoxicity during acute and convalescent SARS-CoV-2 infection (Diao et al., 2020; Mazzoni et al., 2020; H. Y. Zheng et al., 2020a; M. Zheng et al., 2020). Programmed cell death 1 (PD1), T cell immunoglobulin (TIM-3), and mucin-containing protein 3 are exhaustion markers that T cells also exhibit (Shiri et al., 2020). T cell subsets coexpressing Tim-3 and PD1 were identified in patients with severe COVID-19 infections (H. Y. Zheng et al., 2020b). Exhausted CD8+ T cells mainly express PD1, cytotoxic T lymphocyteassociated antigen-4 (CTLA4), T cell immunoglobulin and ITIM domain (TIGIT), and Tim-3. TIGIT expression in CD8+ T cells was shown to correlate with COVID-19 disease progression (H. Y. Zheng et al., 2020b).

However, T cells struggle in clearing SARS-CoV-2 in patients with type 2 diabetes mellitus (T2DM). Hyperglycaemia during T2DM may impair T cell function via the induction of oxidative stress which increases intracellular glucose concentration and mitochondrial proton gradient releasing reactive oxidative species (ROS) (Tong et al., 2021; Xia et al., 2017). Oxidative stress has a clear, detrimental effect on CD8+ T cell responses as it reduces the production of key effector cytokines (Kesarwani et al., 2013). ROS causes structural modifications of the cell receptor signalling proteins (Kesarwani et al., 2013). Another factor that impairs T cell function is glycaemia variability, just like hyperglycaemic, obesity due to adipose and medication such as metformin inhibits the expression of IFN- α in humans via mammalian target of rapamycin (mTOR) complex 1 pathway (Tong et al., 2021; Zatterale et al., 2020).

Metformin also impairs CD4+ cells resulting in a hyperinflammatory response in COVID-19 patients with T2DM (Shoshan-Barmatz et al., 2021).

Co-infection with other diseases such as Tuberculosis (TB) significantly reduces the SARS-CoV-2 -specific immune responses (Petrone et al., 2021; Riou, du Bruyn, et al., 2021). T cells, particularly CD4+ T cells, are part of the immunological response to tuberculosis (Petrone et al., 2021). COVID-19 disease is characterized by lymphopenia but patients with active or latent TB infection can respond to the TB-specific antigens (Tan et al., 2020). In contrast, these patients have a poor immune response to the SARS-CoV-2 infection (Petrone et al., 2021). This observation may be the result of the massive compartmentalization of T cells in infectious foci or by elimination of effector T cells when threatened with high doses of the antigens (Petrone et al., 2021). TB infection impairs the functional capacity and premature functional exhaustion of SARS-CoV-2-specific CD4+ T TB-COVID-19 coinfected patients (Ong et al., 2020; Riou, Du Bruyn, et al., 2021).

Overreaction of the T cell-mediated immune response may lead to complications in severe COVID-19 patients such as pulmonary complications. Pulmonary complications are due to the high expression of CCR4 on the surface of both CD4+ and CD8+ T cells. CCR4 is the lung homing receptor and increased CCR4 expression on T cells promotes a stronger immune response directly in the lungs but this can also result in stronger attraction and activation of T cells in the lungs that may lead to damage to the lungs if not wellregulated (Spoerl et al., 2021). Both CD4+ and CD8+ T cells express significant levels of CD69, CD38 and CD44, in COVID-19 patients with CD4+ T cells also expressing OX40. This is an indication of T cell over activation or overreaction. Furthermore, some investigators have reported a significant increase in the expression of OX40 and 4–1BB in COVID-19 patients who required intensive care unit (ICU) treatment (Y. Zhou et al., 2020) suggesting T cell activation in such patients.

T cell overactivations in COVID-19 patients is characterized by, among other factors, higher level of inflammatory cytokines, other than IFN-γ and the inflammatory cytokine level is related to disease severity. The levels of interleukin (IL) 2R, IL6, IL8, IL10, and TNF- α were significantly higher in severe compared with non-severe patients (Qin et al., 2020). In another study, they reported a lower level of IFN-γ produced by CD4+, CD8+, and natural killer T cells in moderate and severe cases (Chen et al., 2020). Overactivated T cells and increased inflammatory cytokines may enhance the lung's immunopathological damage. (Liu et al., 2020).

4. Conclusion

T cell responses are very critical for immune protection against SARS-CoV-2; however, exacerbated T cell activation may cause T cell exhaustion. There is evidence suggesting that SARS-CoV-2-specific T-cell responses are essential for viral clearance, may prevent infection without seroconversion, provide robust memory, and mediate recognition of two or more viral variants. These T-cell responses are elicited both following natural infection and vaccination. There is some evidence of cross-reactivity with seasonal and or endemic coronaviruses. Despite remarkable progress, there remain many critical questions that need to be resolved about T cell immunity to SARS-CoV-2. Understanding the roles of different subsets of T cells in protection or pathogenesis is crucial for preventing and treating COVID-19. Tcell responses in severely ill patients may be impaired, overactivated, or inappropriate, and further research is required to elucidate this to inform treatment strategies. However, it is unclear whether the strength of the T cell response in the peripheral blood reflects the T cell response

intensity in the respiratory tract and other SARS-CoV-2-infected organs. Further studies should characterize the T cell memory's differentiation state and durability in the tissues or SARS-CoV-2 infected organs. An important application of T cell responses in tissue and SARS-CoV-2-infected organs will be towards the evaluation of the SARS-CoV-2 vaccine and other immunotherapeutic strategies.

Acknowledgements

The authors thank all study participants in the cited papers, the members of the Biological Sciences Departments under the Academy of Medical Sciences for continued support of the research work that produced this paper.

Author contributions

C.M.B: Design, Conceptualisation; Writing, Review. **W.L.M:** Design, conceptualisation, Review. **CK, TN, AK., D.T.M, G.P.B.:** Review.

All authors approved the final manuscript.

Funding

This research received no specific grant from any funding agency in the public, commercial, or notfor-profit sectors. Chikondi Malamba-Banda is PhD student funded by the Bill and Melinda Gates Foundation.

Conflict of interest

The authors declare no conflict of interest.

7. References

Abu-Raddad, L. J., Chemaitelly, H., Ayoub, H. H., Yassine, H. M., Benslimane, F. M., Al Khatib, H. A., Tang, P., Hasan, M. R., Coyle, P., Al Kanaani, Z., Al Kuwari, E., Jeremijenko, A., Kaleeckal, A. H., Latif, A. N., Shaik, R. M., Abdul Rahim, H. F., Nasrallah, G. K., Al Kuwari, M. G., Butt, A. A., … Bertollini, R. (2021). Association of Prior SARS-CoV-2 Infection With Risk of Breakthrough Infection Following mRNA Vaccination in Qatar. JAMA, 326(19), 1930–1939. https://doi.org/10.1001/JAMA.2021.19623

- Akbari, H., Tabrizi, R., Lankarani, K. B., Aria, H., Vakili, S., Asadian, F., Noroozi, S., Keshavarz, P., & Faramarz, S. (2020). The role of cytokine profile and lymphocyte subsets in the severity of coronavirus disease 2019 (COVID-19): A systematic review and meta-analysis. Life Sciences, 258, 118167. https://doi.org/10.1016/J.LFS.2020.118167
- Bates, T. A., McBride, S. K., Winders, B., Schoen, D., Trautmann, L., Curlin, M. E., & Tafesse, F. G. (2022). Antibody Response and Variant Cross-Neutralization After SARS-CoV-2 Breakthrough Infection. JAMA, 327(2), 179–181. https://doi.org/10.1001/JAMA.2021.22898
- Braun, J., Loyal, L., Frentsch, M., Wendisch, D., Georg, P., Kurth, F., Hippenstiel, S., Dingeldey, M., Kruse, B., Fauchere, F., Baysal, E., Mangold, M., Henze, L., Lauster, R., Mall, M. A., Beyer, K., Röhmel, J., Voigt, S., Schmitz, J., … Thiel, A. (2020). SARS-CoV-2-reactive T cells in healthy donors and patients with COVID-19. Nature 2020 587:7833, 587(7833), 270–274. https://doi.org/10.1038/s41586- 020-2598-9

Chen, G., Wu, D., Guo, W., Cao, Y., Huang, D., Wang, H., Wang, T., Zhang, X., Chen, H., Yu, H., Zhang, X., Zhang, M., Wu, S., Song, J., Chen, T., Han, M., Li, S., Luo, X., Zhao, J., & Ning, Q. (2020). Clinical and immunological features of severe and moderate coronavirus disease 2019. The Journal of Clinical Investigation, 130(5), 2620–2629. https://doi.org/10.1172/JCI137244

Chu, H., Zhou, J., Wong, B. H. Y., Li, C., Chan, J. F. W., Cheng, Z. S., Yang, D., Wang, D., Lee, A. C. Y., Li, C., Yeung, M. L., Cai, J. P., Chan, I. H. Y., Ho, W. K., To, K. K. W., Zheng, B. J., Yao, Y., Qin, C., & Yuen, K. Y. (2016). Middle East Respiratory Syndrome Coronavirus Efficiently Infects Human Primary T Lymphocytes and Activates the Extrinsic and Intrinsic Apoptosis Pathways. Journal of Infectious Diseases, 213(6), 904–914. https://doi.org/10.1093/infdis/jiv380

- Diao, B., Wang, C., Tan, Y., Chen, X., Liu, Y., Ning, L., Chen, L., Li, M., Liu, Y., Wang, G., Yuan, Z., Feng, Z., Zhang, Y., Wu, Y., & Chen, Y. (2020). Reduction and Functional Exhaustion of T Cells in Patients with Coronavirus Disease 2019 (COVID-19). Frontiers in Immunology, 11, 827. https://doi.org/10.3389/FIMMU.2020.0082 7/FULL
- Dudakov, J. A., Hanash, A. M., & Van Den Brink, M. R. M. (2015). Interleukin-22: Immunobiology and pathology. Annual Review of Immunology, 33, 747–785. https://doi.org/10.1146/annurev-immunol-032414-112123
- Gadotti, A. C., de Castro Deus, M., Telles, J. P., Wind, R., Goes, M., Garcia Charello Ossoski, R., de Padua, A. M., de Noronha, L., Moreno-Amaral, A., Baena, C. P., & Tuon, F. F. (2020). IFN- γ is an independent risk factor associated with mortality in patients with moderate and severe COVID-19 infection. Virus Research, 289, 198171. https://doi.org/10.1016/J.VIRUSRES.2020. 198171
- García, L. F. (2020). Immune Response, Inflammation, and the Clinical Spectrum of COVID-19. Frontiers in Immunology, 11 (June), $4-8$. https://doi.org/10.3389/fimmu.2020.01441
- Goldberg, Y., Ϯ2, M. M., Bar-On, Y. M., Bodenheimer, O., Freedman, L., Ash, N.,

Alroy-Preis, S., Huppert, A., & &3, R. M. (2021). Protection and waning of natural and hybrid COVID-19 immunity. MedRxiv, 2021.12.04.21267114. https://doi.org/10.1101/2021.12.04.2126711 4

- Grifoni, A., Weiskopf, D., Ramirez, S. I., Mateus, J., Dan, J. M., Moderbacher, C. R., Rawlings, S. A., Sutherland, A., Premkumar, L., Jadi, R. S., Marrama, D., de Silva, A. M., Frazier, A., Carlin, A. F., Greenbaum, J. A., Peters, B., Krammer, F., Smith, D. M., Crotty, S., & Sette, A. (2020). Targets of T Cell Responses to SARS-CoV-2 Coronavirus in Humans with COVID-19 Disease and Unexposed Individuals. Cell, 181(7), 1489-1501.e15. https://doi.org/10.1016/j.cell.2020.05.015
- Harari, A., Dutoit, V., Cellerai, C., Bart, P. A., Du Pasquier, R. A., & Pantaleo, G. (2006). Functional signatures of protective antiviral T-cell immunity in human virus infections. Immunological Reviews, 211(1), 236–254. https://doi.org/10.1111/J.0105- 2896.2006.00395.X
- Jiang, Y., Wei, X., Guan, J., Qin, S., Wang, Z., & Lu, H. (2020). Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19 . The COVID-19 resource centre is hosted on Elsevier Connect , the company ' s public news and information . January.
- Jordan, S. C. (2021). Innate and adaptive immune responses to SARS-CoV-2 in humans: relevance to acquired immunity and vaccine responses. Clinical and Experimental Immunology, 204(3), 310–320. https://doi.org/10.1111/cei.13582
- Juno, J. A., Tan, H. X., Lee, W. S., Reynaldi, A., Kelly, H. G., Wragg, K., Esterbauer, R., Kent, H. E., Batten, C. J., Mordant, F. L.,

Gherardin, N. A., Pymm, P., Dietrich, M. H., Scott, N. E., Tham, W. H., Godfrey, D. I., Subbarao, K., Davenport, M. P., Kent, S. J., & Wheatley, A. K. (2020). Humoral and circulating follicular helper T cell responses in recovered patients with COVID-19. Nature Medicine 2020 26:9, 26(9), 1428– 1434. https://doi.org/10.1038/s41591-020- 0995-0

- Kesarwani, P., Murali, A. K., Al-Khami, A. A., & Mehrotra, S. (2013). Redox regulation of Tcell function: From molecular mechanisms to significance in human health and disease. Antioxidants and Redox Signaling, 18(12), 1497–1534. https://doi.org/10.1089/ars.2011.4073
- Kojima, N., Shrestha, N. K., & Klausner, J. D. (2021). A Systematic Review of the Protective Effect of Prior SARS-CoV-2 Infection on Repeat Infection. Evaluation and the Health Professions, 44(4), 327–332. https://doi.org/10.1177/0163278721104793 2
- Lee, D. C. P., Harker, J. A. E., Tregoning, J. S., Atabani, S. F., Johansson, C., Schwarze, J., & Openshaw, P. J. M. (2010). CD25 + Natural Regulatory T Cells Are Critical in Limiting Innate and Adaptive Immunity and Resolving Disease following Respiratory Syncytial Virus Infection . Journal of Virology, 84(17), 8790–8798. https://doi.org/10.1128/jvi.00796-10
- Li, M. O., & Rudensky, A. Y. (2016). T cell receptor signalling in the control of regulatory T cell differentiation and function. Nature Reviews Immunology 2016 16:4, 16(4), 220–233. https://doi.org/10.1038/nri.2016.26
- Li, T., Qiu, Z., Zhang, L., Han, Y., He, W., Liu, Z., Ma, X., Fan, H., Lu, W., Xie, J., Wang, H., Dang, G., & Wang, A. (2004). Significant Changes of Peripheral T Lymphocyte

Subsets in Patients with Severe Acute Respiratory Syndrome. Journal of Infectious Diseases, 189(4), 648–651. https://doi.org/10.1086/381535

- Liu, L., Xu, L., & Lin, C. (2020). T cell response in patients with COVID-19. Blood Science, 2(3), 76–78. https://doi.org/10.1097/bs9.0000000000000 050
- Loebbermann, J., Thornton, H., Durant, L., Sparwasser, T., Webster, K. E., Sprent, J., Culley, F. J., Johansson, C., & Openshaw, P. J. (2012). Regulatory T cells expressing granzyme B play a critical role in controlling lung inflammation during acute viral infection. Mucosal Immunology, 5(2), 161– 172. https://doi.org/10.1038/mi.2011.62
- Martin, M. D., Kim, M. T., Shan, Q., Sompallae, R., Xue, H. H., Harty, J. T., & Badovinac, V. P. (2015). Phenotypic and Functional Alterations in Circulating Memory CD8 T Cells with Time after Primary Infection. PLoS Pathogens, 11(10), 1–36. https://doi.org/10.1371/journal.ppat.100521 9
- Mazzoni, A., Salvati, L., Maggi, L., Capone, M., Vanni, A., Spinicci, M., Mencarini, J., Caporale, R., Peruzzi, B., Antonelli, A., Trotta, M., Zammarchi, L., Ciani, L., Gori, L., Lazzeri, C., Matucci, A., Vultaggio, A., Rossi, O., Almerigogna, F., … Cosmi, L. (2020). Impaired immune cell cytotoxicity in severe COVID-19 is IL-6 dependent. The Journal of Clinical Investigation, 130(9), 4694–4703.

https://doi.org/10.1172/JCI138554

Moss, P. (2022). The T cell immune response against SARS-CoV-2. Nature Immunology 2022 23:2, 23(2), 186–193. https://doi.org/10.1038/s41590-021-01122 w

- Naqvi, A. A. T., Fatima, K., Mohammad, T., Fatima, U., Singh, I. K., Singh, A., Atif, S. M., Hariprasad, G., Hasan, G. M., & Hassan, M. I. (2020). Insights into SARS-CoV-2 genome, structure, evolution, pathogenesis and therapies: Structural genomics approach. Biochimica et Biophysica Acta - Molecular Basis of Disease, 1866(10), 165878. https://doi.org/10.1016/j.bbadis.2020.16587 8
- Neidleman, J., Luo, X., Frouard, J., Xie, G., Gill, G., Stein, E. S., McGregor, M., Ma, T., George, A. F., Kosters, A., Greene, W. C., Vasquez, J., Ghosn, E., Lee, S., & Roan, N. R. (2020). SARS-CoV-2-Specific T Cells Exhibit Phenotypic Features of Helper Function, Lack of Terminal Differentiation, and High Proliferation Potential. Cell Reports Medicine, $1(6)$. https://doi.org/10.1016/J.XCRM.2020.1000 81
- Newton, R., Priyadharshini, B., & Turka, L. A. (2016). Immunometabolism of regulatory T cells. Nature Immunology 2016 17:6, 17(6), 618–625. https://doi.org/10.1038/ni.3466
- Oberhardt, V., Luxenburger, H., Kemming, J., Schulien, I., Ciminski, K., Giese, S., Csernalabics, B., Lang-Meli, J., Janowska, I., Staniek, J., Wild, K., Basho, K., Marinescu, M. S., Fuchs, J., Topfstedt, F., Janda, A., Sogukpinar, O., Hilger, H., Stete, K., … Hofmann, M. (2021). Rapid and stable mobilization of CD8+ T cells by SARS-CoV-2 mRNA vaccine. Nature 2021 597:7875, 597(7875), 268–273. https://doi.org/10.1038/s41586-021-03841- 4
- Oh, H. L. J., Gan, S. K. E., Bertoletti, A., & Tan, Y. J. (2012). Understanding the T cell immune response in SARS coronavirus infection. Emerging Microbes and

Infections, 1(000), 0. https://doi.org/10.1038/emi.2012.26

- Oh, H. L. J., Gan, S. K. E., Bertoletti, A., & Tan, Y. J. (2019). Understanding the T cell immune response in SARS coronavirus infection. Https://Doi.Org/10.1038/Emi.2012.26, 1. https://doi.org/10.1038/EMI.2012.26
- Ong, C. W. M., Migliori, G. B., Raviglione, M., MacGregor-Skinner, G., Sotgiu, G., Alffenaar, J. W., Tiberi, S., Adlhoch, C., Alonzi, T., Archuleta, S., Brusin, S., Cambau, E., Capobianchi, M. R., Castilletti, C., Centis, R., Cirillo, D. M., D'Ambrosio, L., Delogu, G., Esposito, S. M. R., … Goletti, D. (2020). Epidemic and pandemic viral infections: Impact on tuberculosis and the lung. European Respiratory Journal, 56(4). https://doi.org/10.1183/13993003.01727- 2020
- Or Caspi, Michael J. Smart, R. B. N. (2020). Adaptive immunity to SARS-CoV-2 and COVID-19 Alessandro. Ann Oncol, January, 19–21.
- Papadatou, I., Geropeppa, M., Verrou, K.-M., Tzanoudaki, M., Lagousi, T., Liatsis, E., & Spoulou, V. (2023). SARS-CoV-2 mRNA Dual Immunization Induces Innate Transcriptional Signatures, Establishes T-Cell Memory and Coordinates the Recall Response. Vaccines, 11(1), 103. https://doi.org/10.3390/VACCINES110101 03/S1
- Petrone, L., Petruccioli, E., Vanini, V., Cuzzi, G., Gualano, G., Vittozzi, P., Nicastri, E., Maffongelli, G., Grifoni, A., Sette, A., Ippolito, G., Migliori, G. B., Palmieri, F., & Goletti, D. (2021). Coinfection of tuberculosis and COVID-19 limits the ability to in vitro respond to SARS-CoV-2. International Journal of Infectious Diseases,

113, S82–S87. https://doi.org/10.1016/j.ijid.2021.02.090

- Petrova, G., Ferrante, A., & Gorski, J. (2012). Cross-Reactivity of T Cells and Its Role in the Immune System. Critical Reviews in Immunology, 32(4), 349. https://doi.org/10.1615/CritRevImmunol.v3 2.i4.50
- Qin, C., Zhou, L., Hu, Z., Zhang, S., Yang, S., Tao, Y., Xie, C., Ma, K., Shang, K., Wang, W., & Tian, D. S. (2020). Dysregulation of immune response in patients with coronavirus 2019 (COVID-19) in Wuhan, China. Clinical Infectious Diseases, 71(15), 762–768. https://doi.org/10.1093/cid/ciaa248
- Rahimzadeh, M., & Naderi, N. (2021). Toward an understanding of regulatory T cells in COVID-19: A systematic review. Journal of Medical Virology, 93(7), 4167–4181. https://doi.org/10.1002/jmv.26891
- Riou, C., Du Bruyn, E., Stek, C., Daroowala, R., Goliath, R. T., Abrahams, F., Said-Hartley, Q., Allwood, B. W., Hsiao, M., Wilkinson, K. A., Lindestam Arlehamn, C. S., Sette, A., Wasserman, S., & Wilkinson, R. J. (2021). Profile of SARS-CoV-2-specific CD4 T cell response: relationship with disease severity and impact of HIV-1 and active Mycobacterium tuberculosis co-infection. MedRxiv, 2021.02.16.21251838.
- Riou, C., du Bruyn, E., Stek, C., Daroowala, R., Goliath, R. T., Abrahams, F., Said-Hartley, Q., Allwood, B. W., Hsiao, N. Y., Wilkinson, K. A., Lindestam Arlehamn, C. S., Sette, A., Wasserman, S., & Wilkinson, R. J. (2021). Relationship of SARS-CoV-2– specific CD4 response to COVID-19 severity and impact of HIV-1 and tuberculosis coinfection. Journal of Clinical Investigation, 131(12), 1-15. https://doi.org/10.1172/JCI149125
- Rydyznski Moderbacher, C., Ramirez, S. I., Dan, J. M., Grifoni, A., Hastie, K. M., Weiskopf, D., Belanger, S., Abbott, R. K., Kim, C., Choi, J., Kato, Y., Crotty, E. G., Kim, C., Rawlings, S. A., Mateus, J., Tse, L. P. V., Frazier, A., Baric, R., Peters, B., … Crotty, S. (2020). Antigen-Specific Adaptive Immunity to SARS-CoV-2 in Acute COVID-19 and Associations with Age and Disease Severity. Cell, 183(4), 996- 1012.e19. https://doi.org/10.1016/j.cell.2020.09.038
- Sahin, U., Muik, A., Vogler, I., Derhovanessian, E., Kranz, L. M., Vormehr, M., Quandt, J., Bidmon, N., Ulges, A., Baum, A., Pascal, K. E., Maurus, D., Brachtendorf, S., Lörks, V., Sikorski, J., Koch, P., Hilker, R., Becker, D., Eller, A. K., … Türeci, Ö. (2021). BNT162b2 vaccine induces neutralizing antibodies and poly-specific T cells in humans. Nature 2021 595:7868, 595(7868), 572–577. https://doi.org/10.1038/s41586- 021-03653-6
- Schmetterer, K. G., Neunkirchner, A., & Pickl, W. F. (2012). Naturally occurring regulatory T cells: markers, mechanisms, and manipulation. FASEB Journal: Official Publication of the Federation of American Societies for Experimental Biology, 26(6), 2253–2276. https://doi.org/10.1096/FJ.11- 193672
- Scudellari, Megan. (2021). How the coronavirus infects our cells. Nature , 595(July).
- Seder, R. A., Darrah, P. A., & Roederer, M. (2008). T-cell quality in memory and protection: Implications for vaccine design. Nature Reviews Immunology, 8(4), 247–258. https://doi.org/10.1038/nri2274
- Sekine, T., Perez-Potti, A., Rivera-Ballesteros, O., Strålin, K., Gorin, J. B., Olsson, A., Llewellyn-Lacey, S., Kamal, H., Bogdanovic, G., Muschiol, S., Wullimann,

D. J., Kammann, T., Emgård, J., Parrot, T., Folkesson, E., Akber, M., Berglin, L., Bergsten, H., Brighenti, S., … Buggert, M. (2020a). Robust T Cell Immunity in Convalescent Individuals with Asymptomatic or Mild COVID-19. Cell, 183(1), 158-168.e14. https://doi.org/10.1016/J.CELL.2020.08.01 7

- Sekine, T., Perez-Potti, A., Rivera-Ballesteros, O., Strålin, K., Gorin, J. B., Olsson, A., Llewellyn-Lacey, S., Kamal, H., Bogdanovic, G., Muschiol, S., Wullimann, D. J., Kammann, T., Emgård, J., Parrot, T., Folkesson, E., Akber, M., Berglin, L., Bergsten, H., Brighenti, S., … Buggert, M. (2020b). Robust T Cell Immunity in Convalescent Individuals with Asymptomatic or Mild COVID-19. Cell, 183(1), 158-168.e14. https://doi.org/10.1016/J.CELL.2020.08.01 7
- Sette, A., & Crotty, S. (2021). Adaptive immunity to SARS-CoV-2 and COVID-19. Cell, 184(4), 861. https://doi.org/10.1016/J.CELL.2021.01.00 7
- Shin, H. S., Kim, Y., Kim, G., Lee, J. Y., Jeong, I., Joh, J. S., Kim, H., Chang, E., Sim, S. Y., Park, J. S., & Lim, D. G. (2019). Immune Responses to Middle East Respiratory Syndrome Coronavirus During the Acute and Convalescent Phases of Human Infection. Clinical Infectious Diseases, 68(6), 984–992. https://doi.org/10.1093/CID/CIY595
- Shiri, P., Eslami, N., Shamekh, A., & Entezarimaleki, T. (2020). Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted

on Elsevier Connect, the company ' s public news and information . January.

- Shoshan-Barmatz, V., Anand, U., Nahon-Crystal, E., Di Carlo, M., & Shteinfer-Kuzmine, A. (2021). Adverse Effects of Metformin From Diabetes to COVID-19, Cancer, Neurodegenerative Diseases, and Aging: Is VDAC1 a Common Target? Frontiers in Physiology, 12 (October). https://doi.org/10.3389/fphys.2021.730048
- Skelly, D. T., Harding, A. C., Gilbert-Jaramillo, J., Knight, M. L., Longet, S., Brown, A., Adele, S., Adland, E., Brown, H., Chinnakannan, S., Donnison, T., Ali, M., Rongkard, P., Pace, M., Zacharopoulou, P., Robinson, N., Csala, A., De Lara, C., Hutchings, C. L., … James, W. S. (2021). Two doses of SARS-CoV-2 vaccination induce robust immune responses to emerging SARS-CoV-2 variants of concern. Nature Communications 2021 12:1, 12(1), 1–12. https://doi.org/10.1038/s41467-021-25167- 5
- Spoerl, S., Kremer, A. N., Aigner, M., Eisenhauer, N., Koch, P., Meretuk, L., Löffler, P., Tenbusch, M., Maier, C., Überla, K., Heinzerling, L., Frey, B., Lutzny-Geier, G., Winkler, T. H., Krönke, G., Vetter, M., Bruns, H., Neurath, M. F., Mackensen, A., … Völkl, S. (2021). Upregulation of CCR4 in activated CD8+ T cells indicates enhanced lung homing in patients with severe acute SARS-CoV-2 infection. European Journal of Immunology, 51(6), 1436–1448.

https://doi.org/10.1002/EJI.202049135

Steiner, S., Sotzny, F., Bauer, S., Na, I.-K., Schmueck-Henneresse, M., Corman, V. M., Schwarz, T., Drosten, C., Wendering, D. J., Behrends, U., Volk, H.-D., Scheibenbogen, C., & Hanitsch, L. G. (2020). HCoV- and SARS-CoV-2 Cross-Reactive T Cells in CVID Patients. Frontiers in Immunology, 0, 3347. https://doi.org/10.3389/FIMMU.2020.6079 18

- Suvas, S., Azkur, A. K., Kim, B. S., Kumaraguru, U., & Rouse, B. T. (2004). CD4 + CD25 + Regulatory T Cells Control the Severity of Viral Immunoinflammatory Lesions . The Journal of Immunology, 172(7), 4123–4132. https://doi.org/10.4049/jimmunol.172.7.412 3
- Tan, L., Wang, Q., Zhang, D., Ding, J., Huang, Q., Tang, Y. Q., Wang, Q., & Miao, H. (2020). Lymphopenia predicts disease severity of COVID-19: a descriptive and predictive study. Signal Transduction and Targeted Therapy, 5(1), 16–18. https://doi.org/10.1038/s41392-020-0148-4
- Tincati, C., Cannizzo, E. S., Giacomelli, M., Badolato, R., d'Arminio Monforte, A., & Marchetti, G. (2020). Heightened Circulating Interferon-Inducible Chemokines, and Activated Pro-Cytolytic Th1-Cell Phenotype Features Covid-19 Aggravation in the Second Week of Illness. Frontiers in Immunology, 11, 2672. https://doi.org/10.3389/FIMMU.2020.5809 87/BIBTEX
- Tong, Z. W. M., Grant, E., Gras, S., Wu, M., Smith, C., Barrett, H. L., Gallo, L. A., & Short, K. R. (2021). The role of T-cell immunity in COVID-19 severity amongst people living with type II diabetes. FEBS Journal, 288(17), 5042–5054. https://doi.org/10.1111/febs.16105
- Toor, S. M., Saleh, R., Sasidharan Nair, V., Taha, R. Z., & Elkord, E. (2021). T-cell responses and therapies against SARS-CoV-2 infection. Immunology, 162(1), 30–43. https://doi.org/10.1111/imm.13262
- van der Veeken, J., Gonzalez, A. J., Cho, H., Arvey, A., Hemmers, S., Leslie, C. S., & Rudensky, A. Y. (2016). Memory of Inflammation in Regulatory T Cells. Cell, 166(4), 977–990. https://doi.org/10.1016/J.CELL.2016.07.00 6
- Walls, A. C., Park, Y. J., Tortorici, M. A., Wall, A., McGuire, A. T., & Veesler, D. (2020). Structure, Function, and Antigenicity of the SARS-CoV-2 Spike Glycoprotein. Cell, 181(2), 281-292.e6. https://doi.org/10.1016/j.cell.2020.02.058
- Wang, H., Wang, Z., Cao, W., Wu, Q., Yuan, Y., & Zhang, X. (2021). Regulatory T cells in COVID-19. Aging and Disease, 12(7), 1545–1553. https://doi.org/10.14336/AD.2021.0709
- Wang, M. Y., Zhao, R., Gao, L. J., Gao, X. F., Wang, D. P., & Cao, J. M. (2020). SARS-CoV-2: Structure, Biology, and Structure-Based Therapeutics Development. Frontiers in Cellular and Infection Microbiology, 10(November), 1–17. https://doi.org/10.3389/fcimb.2020.587269
- Wang, W., Su, B., Pang, L., Qiao, L., Feng, Y., Ouyang, Y., Guo, X., Shi, H., Wei, F., Su, X., Yin, J., Jin, R., & Chen, D. (2020). Highdimensional immune profiling by mass cytometry revealed immunosuppression and dysfunction of immunity in COVID-19 patients. Cellular and Molecular Immunology, 17(6), 650–652. https://doi.org/10.1038/s41423-020-0447-2
- Wang, Y., Sibaii, F., Lee, K., J. Gill, M., & L. Hatch, J. (2021). NOTE: This preprint reports new research that has not been certified by peer review and should not be used to guide clinical practice. 1. MedRxiv, 1(165), 1–13.
- Weiskopf, D., Schmitz, K. S., Raadsen, M. P., Grifoni, A., Okba, N. M. A., Endeman, H., van den Akker, J. P. C., Molenkamp, R., Koopmans, M. P. G., van Gorp, E. C. M., Haagmans, B. L., de Swart, R. L., Sette, A., & de Vries, R. D. (2020a). Phenotype and kinetics of SARS-CoV-2-specific T cells in COVID-19 patients with acute respiratory distress syndrome. Science Immunology, $5(48)$, $1-11$. https://doi.org/10.1126/SCIIMMUNOL.AB D2071
- Weiskopf, D., Schmitz, K. S., Raadsen, M. P., Grifoni, A., Okba, N. M. A., Endeman, H., van den Akker, J. P. C., Molenkamp, R., Koopmans, M. P. G., van Gorp, E. C. M., Haagmans, B. L., de Swart, R. L., Sette, A., & de Vries, R. D. (2020b). Phenotype and kinetics of SARS-CoV-2-specific T cells in COVID-19 patients with acute respiratory distress syndrome. Science Immunology, 5(48), 2071. https://doi.org/10.1126/SCIIMMUNOL.AB D2071/SUPPL_FILE/ABD2071_TABLE_ S1.XLSX
- Weissman, I. L. (2000). Stem cells: Units of development, units of regeneration, and units in evolution. Cell, 100(1), 157–168. https://doi.org/10.1016/S0092- 8674(00)81692-X
- Wrapp, D., Wang, N., Corbett, K. S., Goldsmith, J. A., Hsieh, C. L., Abiona, O., Graham, B. S., & McLellan, J. S. (2020). Cryo-EM structure of the 2019-nCoV spike in the prefusion conformation. Science, 367(6483), 1260–1263. https://doi.org/10.1126/science.aax0902
- Wu, F., Zhao, S., Yu, B., Chen, Y. M., Wang, W., Song, Z. G., Hu, Y., Tao, Z. W., Tian, J. H., Pei, Y. Y., Yuan, M. L., Zhang, Y. L., Dai, F. H., Liu, Y., Wang, Q. M., Zheng, J. J., Xu, L., Holmes, E. C., & Zhang, Y. Z. (2020). A

new coronavirus associated with human respiratory disease in China. Nature, 579(7798), 265–269. https://doi.org/10.1038/s41586-020-2008-3

- Xia, C., Rao, X., & Zhong, J. (2017). Role of T Lymphocytes in Type 2 Diabetes and Diabetes-Associated Inflammation. Journal of Diabetes Research, 2017. https://doi.org/10.1155/2017/6494795
- Zatterale, F., Longo, M., Naderi, J., Raciti, G. A., Desiderio, A., Miele, C., & Beguinot, F. (2020). Chronic Adipose Tissue Inflammation Linking Obesity to Insulin Resistance and Type 2 Diabetes. Frontiers in Physiology, 10 (January), $1-20$. https://doi.org/10.3389/fphys.2019.01607
- Zeng, C., Evans, J. P., King, T., Zheng, Y. M., Oltz, E. M., Whelan, S. P. J., Saif, L. J., Peeples, M. E., & Liu, S. L. (2022). SARS-CoV-2 spreads through cell-to-cell transmission. Proceedings of the National Academy of Sciences of the United States of America, 119(1). https://doi.org/10.1073/PNAS.2111400119/ SUPPL_FILE/PNAS.2111400119.SAPP.P
- Zheng, H. Y., Zhang, M., Yang, C. X., Zhang, N., Wang, X. C., Yang, X. P., Dong, X. Q., & Zheng, Y. T. (2020a). Elevated exhaustion levels and reduced functional diversity of T cells in peripheral blood may predict severe progression in COVID-19 patients. Cellular & Molecular Immunology, 17(5), 541–543. https://doi.org/10.1038/S41423-020-0401-3
- Zheng, H. Y., Zhang, M., Yang, C. X., Zhang, N., Wang, X. C., Yang, X. P., Dong, X. Q., &

Zheng, Y. T. (2020b). Elevated exhaustion levels and reduced functional diversity of T cells in peripheral blood may predict severe progression in COVID-19 patients. Cellular $&$ Molecular Immunology, 17(5), 541–543. https://doi.org/10.1038/S41423-020-0401-3

- Zheng, M., Gao, Y., Wang, G., Song, G., Liu, S., Sun, D., Xu, Y., & Tian, Z. (2020). Functional exhaustion of antiviral lymphocytes in COVID-19 patients. Cellular and Molecular Immunology, 17(5), 533. https://doi.org/10.1038/S41423-020- 0402-2
- Zhou, P., Yang, X. Lou, Wang, X. G., Hu, B., Zhang, L., Zhang, W., Si, H. R., Zhu, Y., Li, B., Huang, C. L., Chen, H. D., Chen, J., Luo, Y., Guo, H., Jiang, R. Di, Liu, M. Q., Chen, Y., Shen, X. R., Wang, X., … Shi, Z. L. (2020). A pneumonia outbreak associated with a new coronavirus of probable bat origin. Nature, 579(7798), 270–273. https://doi.org/10.1038/s41586-020-2012-7
- Zhou, Y., Fu, B., Zheng, X., Wang, D., Zhao, C., qi, Y., Sun, R., Tian, Z., Xu, X., & Wei, H. (2020). Aberrant pathogenic GM-CSF+ T cells and inflammatory CD14+CD16+ monocytes in severe pulmonary syndrome patients of a new coronavirus. BioRxiv, 2020.02.12.945576.

https://doi.org/10.1101/2020.02.12.945576

DF